

Identifying patterns and drivers of coral diversity in the Central Indo-Pacific marine biodiversity hotspot

Morana Mihaljević, Chelsea Korpany, Willem Renema, Kevin Welsh and John M. Pandolfi

Abstract.—Biodiversity hotspots are increasingly recognized as areas of high taxonomic and functional diversity. These hotspots are dynamic and shift geographically over time in response to environmental change. To identify drivers of the origin, evolution, and persistence of diversity hotspots, we investigated the diversity patterns of reef-building corals (Scleractinia) in the Central Indo-Pacific, a marine biodiversity hotspot for the last 25 Myr. We used the scleractinian fossil record (based on literature and a newly acquired fossil collection) to examine the taxonomic and functional diversity of corals from the Eocene to Pliocene. Our data identify potential drivers of coral diversity through time (and space) in the Central Indo-Pacific by constraining the timing of taxonomic turnover events and correlating them with known environmental changes. Increases in taxonomic diversity, high origination rates, and changes in abundance of functional character states indicate that the origin of the Central Indo-Pacific hotspot occurred during the Oligocene, initially through proliferation of pre-existing taxa and then by origination of new taxa. In contrast to taxonomic diversity, overall functional diversity of Central Indo-Pacific reef-building corals remained constant from the Eocene to the Pliocene. Our results identify global sea level as a main driver of diversity increase that, together with local tectonics, regulates availability of suitable habitats. Moreover, marine biodiversity hotspots develop from both the accumulation of taxa from older biodiversity hotspots and origination of new taxa. Our study demonstrates the utility of a combined literature-based and new collection approach for gaining new insights into the timing, drivers, and development of tropical biodiversity hotspots.

Morana Mihaljević and Kevin Welsh. School of Earth and Environmental Sciences, University of Queensland, Brisbane, Queensland 4072, Australia. E-mail: m.mihaljevic@uq.edu.au, k.welsh1@uq.edu.au.

Chelsea Korpany and John M. Pandolfi. Australian Research Council Centre of Excellence for Coral Reef Studies, School of Biological Sciences, University of Queensland, Brisbane, Queensland 4072, Australia. E-mail: c.korpany@uq.edu.au, j.pandolfi@uq.edu.au.

Willem Renema. Naturalis Biodiversity Center, 2333 CR Leiden, Netherlands. E-mail: willem.renema@naturalis.nl.

Accepted: 3 January 2017

Data available from the Dryad Digital Repository: <http://dx.doi.org/10.5061/dryad.3k5v6>

Introduction

Biodiversity hotspots are areas with exceptionally high regional taxonomic diversity that arise from spatial differences in speciation, immigration, and extinction rates (MacArthur and Wilson 1967; Russel 1998; Bellwood et al. 2012). However, the definition of biodiversity has broadened recently from a measure of species richness to a more comprehensive concept that involves not only the number and abundance of species present but also their functional traits (Moore 2013; Stuart-Smith et al. 2013). Functional traits (e.g., morphological, physiological, behavioral) determine the interactions of organisms with their environment and with each other (Violle et al. 2007; Díaz et al. 2013; Kraft et al. 2015). Therefore, it

is crucial to explore both the abundance of species and their functional traits in order to understand regional biodiversity (Luck et al. 2009; Devictor et al. 2010).

Biogeographic patterns vary through time and space as a consequence of the interplay among origination, extinction, and migration. By constraining the timing of taxonomic turnover events and correlating them with known environmental changes, it is possible to identify drivers of diversity through time and space. For example, systematic analyses of the fossil record of the tropical marine fauna and coral reef systems of the Caribbean region have revealed patterns of diversity and species distribution through geological time (Jackson et al. 1996; Budd 2000; Johnson et al. 2008;

Klaus et al. 2011). During the Cenozoic, a constant increase in generic coral diversity in the Caribbean from 50 Ma was interrupted in the early Miocene at about ~23 Ma; this interruption corresponds to regional changes in water quality that led to range restrictions and regional extinctions (Stehli and Wells 1971; Edinger and Risk 1994, 1995; Johnson et al. 2009). The Neogene closure of the Central American Isthmus also triggered changes in water quality and other environmental factors (e.g., productivity) (O'Dea and Collins 2013; Leigh et al. 2014). These changes had profound impacts on coral reefs and the tropical biodiversity they supported throughout the Neogene (Jackson et al. 1996; Marko et al. 2015; O'Dea et al. 2016). In comparison to the work in the Caribbean basin, relatively little is known about the development of the modern Central Indo-Pacific marine diversity hotspot, including the history of origination–extinction events and their relationship to environmental changes over time. Here, we use the Central Indo-Pacific marine diversity hotspot to investigate patterns and characteristics of hotspot development through time.

At least since the early Miocene (~23–15.97 Ma), the Central Indo-Pacific (20°S to 20°N, 90° E to 180°E) has persisted as a hotspot of global marine diversity (Wilson and Rosen 1998; Renema et al. 2008; Lohman et al. 2011). However, at least two earlier marine diversity hotspots, the West Tethyan (middle Eocene and older) and the Arabian (late Eocene–early Miocene), are recognized during the Cenozoic (65 Ma–present) along the northern shores of the Paleo-Tethys ocean (Renema et al. 2008). These hotspots are associated with a high diversity of marine taxa such as foraminifera (Renema 2007; Renema et al. 2008), mollusks (Kay 1996), mangroves (Ellison et al. 1999; Morley 2000), and corals (Wilson and Rosen 1998; Wallace and Rosen 2006). Areas with a high diversity of marine taxa shifted from the West Tethyan region across the Middle East to the Central Indo-Pacific, where the modern high-diversity region was established by the early Miocene.

The establishment of the high diversity in the Central Indo-Pacific appears to have been facilitated by tectonic events during the

Cenozoic (Renema et al. 2008). In particular, substantial expansion of the shallow-marine habitats suitable for coral reef development resulted from the opening of the South China Sea (~45–17 Ma) and the collision of Australia with the Pacific arcs and the Southeast Asian margin (starting ~20–25 Ma) (Hall 1996, 2002, 2009; Renema et al. 2008; Hall et al. 2011). The reported increase in shallow-marine habitats from the middle Eocene to the early Miocene (Mihaljević et al. 2014) coincided with abrupt increases in coral abundance, measured as area of coral-dominated carbonate platforms (Wilson 2008). However, it is unclear whether the abrupt increase of coral abundance was accompanied by an increase in coral diversity (Wilson and Rosen 1998).

Knowledge of the pre–early Miocene coral fossil record is sparse, so much so that Wilson and Rosen (1998) refer to the paucity of Paleogene fossil corals as the “Paleogene gap.” However, recent discovery of abundant and diverse Oligocene coral assemblages from Sabah, Malaysia (McMonagle et al. 2011; McMonagle 2012), call into question the timing of the origin of the Central Indo-Pacific biodiversity hotspot. Most genera of extant reef-building corals in the Central Indo-Pacific were present between the early–middle Miocene and the Pliocene (Umbgrove 1946; Wilson and Rosen 1998; Santodomingo et al. 2015a, 2016). Detailed analysis of the Indonesian coral fossil record shows a faunal turnover in the early Miocene (Burdigalian) that resulted in an increase in coral diversity followed by relatively high diversity and limited turnover from the Miocene to the early Pleistocene (Johnson et al. 2015; Santodomingo et al. 2016). However, Bromfield and Pandolfi (2011) recorded two intervals of heightened taxonomic turnover between the middle Miocene and the Pleistocene in the Central Indo-Pacific (Indonesia, Papua New Guinea, and Fiji). The first interval corresponded with an origination event that resulted in an increase in coral generic richness in the middle Miocene. This origination event was followed by a gradual decrease in richness throughout the late Miocene and Pliocene. Thus, given the existing disparities among regional analyses, robust estimates of coral diversity in the region

and the timing of the origin of the Central Indo-Pacific hotspot can only be obtained through the acquisition of new fossil collections from diverse environments across the Central Indo-Pacific throughout the Cenozoic era (Johnson et al. 2015; Wilson 2015).

Here, we use the scleractinian fossil record of the Central Indo-Pacific to investigate the taxonomic and functional diversity and community composition of corals from the Eocene to the Pliocene. By combining new records of Oligocene–early Miocene (Aquitainian) fossil corals from Malaysia and the Philippines with records previously reported from the Central Indo-Pacific region, we aim to refine the timing of the origin of the modern Central Indo-Pacific biodiversity hotspot, as defined by broad patterns in taxonomic and functional diversities of reef corals, and to identify potential drivers of diversity through time. In doing so, we provide a detailed record of temporal ranges of fossil corals in the region, and we estimate Neogene origination and extinction rates of Indo-Pacific coral genera and coral functional traits across 55 Myr. These data on the Central Indo-Pacific fossil record represent our most detailed view of the development and persistence of a biodiversity hotspot to date and yield important new ecological insights, particularly with regard to origination and extinction during the initiation and buildup of regional diversity. This example is not only useful in increasing our knowledge of hotspot dynamics, but also in providing a framework from which to examine high-diversity regions and understand their vulnerability and or adaptability to future environmental changes.

Materials and Methods

Study Sites and Sampling.—Fossil corals ($N = 1606$ coral fragments) were collected from three areas within the Central Indo-Pacific: Sarawak in Malaysia (Melinau and Subis [Tangap] Limestones) and the islands of Negros (Teankalan/Binaguiohan Limestone) and Cebu (Calagasan Formation and Butong Limestone Formation) in the Philippines (Fig. 1). Stratigraphic ranges of geologic sections from the study sites were determined using occurrences of large benthic foraminifera (LBF)

zonation (Ta–Tf) (Lunt and Allan 2004; Renema 2007; Lunt and Renema 2014). The LBF zonation is a common biostratigraphical tool in Cenozoic shallow-marine tropical deposits that allows dating accuracy of 1–3 million years (Renema 2007; Lunt and Renema 2014). Formations were sampled based upon their prior reported ages (Oligocene and early Miocene), previous site descriptions and locality information (Barnes et al. 1958; Adams and Haak 1962; Adams 1965; Jurgan and Domingo 1989; Porth and von Daniels 1989; Wannier 2009; Aurelio and Peña 2010; Mihaljević et al. 2014), and field observations regarding the presence, abundance, and quality of preservation of fossil corals. Both limestone and shale formations/lithologies were sampled in order to capture the range of sedimentary environments and coral reef habitats.

Fossil corals were collected in stratigraphic sequence from outcrops at each study site (14 localities within five geological formations) (Fig. 1, Table 1, Supplementary Table 1). Corals in the Subis Limestone were collected from two different sections from the S&Y Quarry in Batu Niah: (1) a single bed at least 15 m thick with coral fragments embedded in fine matrix, and (2) a 10-m-thick section (five beds) characterized by corals and large benthic foraminifera associated with subordinate coralline algae and mollusk fragments. The general lithology of the deposits are coral/algal grainstone to boundstone forming part of an unattached platform (for details, see Mihaljević et al. 2014). The Melinau Limestone Formation locality consisted of a 13 m³ isolated limestone block/olistolith of coral/algal boundstone and foraminifera packstone derived from a reefal body set in the younger Setap Shale Formation (Wannier 2009) with excellent in situ preservation of corals (for details, see Mihaljević et al. 2014). Corals in the Teankalan/Binaguiohan Limestone locality were collected along a single, highly recrystallized, coral-rich bed of up to 15 m in thickness exposed on the road going from Candoni south into the valley. In general, the Teankalan/Binaguiohan Limestone is characterized by pink or cream to white, thick, coarse-grained micritic bioarenite beds (sometimes brecciated) that grade into irregular lenses of micritic coral rudites.

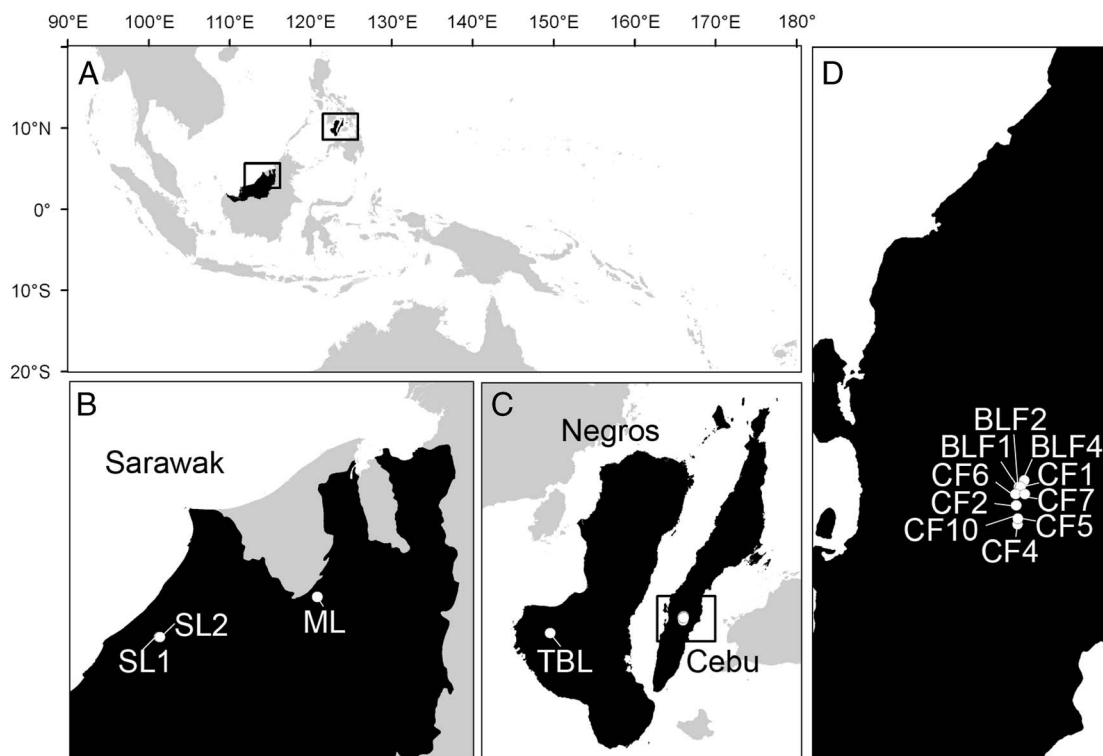


FIGURE 1. Map of study sites where new fossil coral collections were made. A, Map of the Central Indo-Pacific region with Sarawak, Negros and Cebu marked in black. B, Sarawak with two localities from the Subis Limestone (SL1 and SL2) and one locality from the Melinau Limestone (ML). C, Close up of islands of Negros and Cebu in the Philippines. TBL locality from Negros is indicated. Black rectangle and white dot in the southern part of Cebu mark general position of the localities shown in D. D, Map of the studied localities on Cebu: seven localities from the Calagasan Formation (CF1, CF2, CF4, CF5, CF6, CF7, and CF10) and three localities from the Butong Limestone Formation (BLF1, BLF2, and BLF4).

In addition to corals, large benthic foraminifera, mollusks, and coralline algae are also locally abundant (Jurgan and Domingo 1989; Porth and von Daniels 1989; Aurelio and Peña 2010). Fossil coral collection in both the Calagasan (seven localities) and Butong Limestone Formation (three locations) occurred opportunistically along road cuts in coral-rich horizons while driving from Argao to Delaguete on the inland/mountain dirt roads. The Butong Limestone Formation (which is the southern equivalent of the Cebu Limestone) consists of bedded packstones and biorudites characterized by the presence of large benthic foraminifera and corals and high local abundances of branching corals. The Butong Limestone Formation overlies and interfingers with the Calagasan Formation (Porth and von Daniels 1989; Wilson 2002; Aurelio and Peña 2010). The Calagasan Formation is mostly

sandstone and mudstone with sporadic limestone beds. The limestone beds, set in brown to dark-greenish carbonaceous shale, are rich with coral and large benthic foraminifera. Corals are found in situ within the framestone of the limestone beds and also scattered in the shale below. All localities within the same formation, except the two Subis localities, can be grouped into a single faunule (an association of animal fossils in a single stratum or succession of strata of limited thickness; sensu Jackson et al. 1996). Therefore, the collected fossil corals have been grouped into six faunules: Subis 1, Subis 2, Melinau, Teankalan/Binaguiohan, Calagasan, and Butong.

Sampling efforts were standardized by setting time limits to coral collection within a 10-m-thick section. The time limits differed among sites with different lithologies (1h/section for shale; 3h/section for massive limestone) in

TABLE 1. Age, depositional setting, and lithology of geological formations studied.

Geological Formation	Age	Tectonic setting	Depositional setting	Lithology	Biota	References
Calagasan Formation (Cebu)	Late Oligocene		Coral-rich limestone lenses within a shale, potentially patchy reefs	Conglomerate, sandstone and shale with coal and limestone interbeds	Coral and large benthic foraminifera common and locally abundant in limestone interbeds	Porth et al. 1989; Aurelio and Peña 2010
Butong Limestone Formation (Cebu)	Late Oligocene–early Miocene	Backarc setting? Oblique convergence	Isolated low-relief buildups	Bedded biomicrites, biorudites, and bioarenites	Common to abundant <i>Lepidocyclus</i> and other larger foraminifera; locally branching and finger corals abundant	Porth et al. 1989; Wilson 2002; Aurelio and Peña 2010
Trankalan/Binaguiohan Limestone (Negros)	Late Oligocene–early Miocene	Unclear	Carbonate platform with localized patch reefs	Rudstones, bioclastic packstones, and limestone breccias	Coralline algae, green algae, large benthic foraminifera, corals (head and branching corals), echinoderms, and mollusks	Jurgan and Domingo, 1989; Porth et al. 1989; Wilson 2002; Aurelio and Peña 2010
Melinau Limestone (Sawarwak)	Eocene–early Miocene	Isolated carbonate platform, basin margin	Carbonate platform, some marginal deposits, coral patch reefs only noted in Miocene	Packstones, grainstones, some redeposited beds, some dolomitization	Larger benthic foraminifera dominate, coralline algae; Corals in large benthic foraminifera zone Te	Adams and Haak 1962; Adams 1965; Wilson 2002; Wannier 2009; Mihaljević et al. 2014
Subis Limestone (Sarawak)	Early Miocene	Baram Balabac Basin margin	Isolated carbonate platform formed on paleohigh	Foraminiferal–algal wackestone to packstone, floatstone, grainstone	Large benthic foraminifera, coralline algae, and corals	Adams 1965; Wilson 2002; Mihaljević et al. 2014

order to account for the more difficult, time-consuming extraction of samples from highly indurated massive limestones. The coral specimens extracted from massive limestones were cut with a rock saw prior to analysis to expose diagnostic characters and to create thin sections.

Taxonomy.—All collected fossil coral specimens were identified to genus (M. Mihaljević unpublished data). The Coral ID interactive key (Veron and Stafford-Smith 2002) and recent changes in taxonomic delineations (e.g., Huang et al. 2014) were used to identify extant coral taxa. The paleontological literature (Supplementary Table 2), published monographs (Leloux and Renema 2007; Bromfield 2013), and a reference collection from the Queensland Museum were used to identify all extinct fossil genera.

To accurately capture taxonomic turnover and to estimate coral diversity through time, the data from our new collection were combined with records of fossil corals derived from the Paleobiology Database (PBDB; Supplementary Table 2) in April 2016 and supplemented with recent literature published through April 2016 (Leloux and Renema 2007; McMonagle 2012; Bromfield 2013; Novak et al. 2013; Kusworo et al. 2015; Santodomingo et al. 2015a,b). To search and download relevant fossil coral records from the PBDB, the following search criteria were used: “Scleractinia”; “Paleogene to Quaternary.” From this global data set, we extracted records from the Central Indo-Pacific region (20°S to 20°N, 90°E to 180°E). The extracted PBDB Central Indo-Pacific coral data set was reviewed to address limitations inherent with the use of published records from the PBDB, such as the need to account for collector and facies biases as well as inconsistencies with current nomenclature. Therefore, prior to the analysis we: (1) cross-checked taxonomic nomenclature of each genus with the current literature and the World Register of Marine Species (WoRMS Editorial Board 2016), updating where necessary; (2) standardized stratigraphic nomenclature based on the International Commission on Stratigraphy (Cohen et al. 2013); and (3) excluded from analysis all records with poorly defined stratigraphy (site age not at the appropriate scale of resolution; e.g., Neogene).

Fossil data derived from the PBDB, recent literature, and this study are listed in Supplementary Table 2.

Estimating True Stratigraphic Range.—The first-appearance datum and last-appearance datum (FAD and LAD, respectively) of a taxon define the observed end points of its temporal range. Estimating the true range of a taxon is fundamental for any evolutionary analysis. Due to sampling and preservation biases, the observed range of a fossil taxon is almost certainly a truncated version of its true stratigraphic range (Strauss and Sadler 1989). To express the uncertainty of observed taxon ranges, confidence intervals for FADs and LADs were estimated as the average gap length between occurrences within their observed range (Strauss and Sadler 1989; Marshall 1990; Hammer and Harper 2005). The observed range of a taxon was obtained by compiling temporal ranges of the newly collected specimens, specimens reported from the PBDB, and the recent literature.

Taxonomic Diversity.—Diversity was estimated using two independent methods: mean standing diversity (MSD) (Sepkoski 1975; Foote 2000; Hammer 2003) and the Chao 2 index (Chao 1984, 1987). Although these two methods are similar to one another, they have intrinsic differences that cause them to underestimate (MSD) and overestimate (Chao2) diversity (e.g., Foggo et al. 2003; Tammekänd et al. 2010). By using both methods, we hoped to gain an understanding of the range and variability of diversity measurements. However, for most of the discussion we will refer to the more conservative diversity estimate: MSD.

MSD for a specific time interval was calculated by differentially weighting taxa based on their occurrence through time (Sepkoski 1975; Foote 2000; Hammer 2003). For instance, taxa that range throughout a time interval are weighted as 1 unit, taxa that have either a first or last occurrence in that specific time interval as 1/2 unit, and taxa that occur within a single time interval as 1/3 unit. In our study, the time intervals are equivalent to the time stages defined by the International Commission on Stratigraphy (Cohen et al. 2013). Using this weighting method minimizes distortions of the fossil record caused by variation in

preservation and stage length; it is commonly used (e.g., Smith 2001; Jaramillo 2002; Klug et al. 2010) because it assumes that a taxon is present in all samples that lie between its FAD and LAD. In this way, the potential for sampling bias on standing diversity is reduced.

The Chao 2 index is often used in ecology for analyzing capture–recapture experiments, but it has also been shown to be a good estimator of diversity (Colwell and Coddington 1994). This index takes into account that not all taxa are equally common in an environment and therefore are not equally represented in fossil samples. The Chao 2 index and its 95% confidence intervals were calculated using the EstimateS software (Colwell 2013). To detect the adequacy of our sampling efforts, a sampling curve was generated by plotting the number of collected fossil corals against the number of genera. The sampling curve was generated using R, Version 2.15.2 software (R Development Core Team 2012). Finally, we investigated the relationship between coral diversity and area of coral-dominated carbonates (estimated from Wilson 2008: Fig. 2) through time using linear regression.

Taxonomic Turnover.—To investigate taxonomic turnover of corals in the Central Indo-Pacific, we calculated origination and extinction rates. Because we are studying diversity dynamics within a limited geographical region (Central Indo-Pacific) and not globally, the term “origination” refers to the addition of taxa new to the region through both origination and immigration. Similarly, the term “extinction” represents the regional loss of taxa through extinction and/or emigration. Origination/immigration and extinction/emigration rates, herein just referred to as origination and extinction rates, were calculated by dividing the number of extinctions/emigrations (or originations/immigrations) within the time interval (geologic stage) by the estimated MSD. We then divided this value by the length of the time interval to determine an extinction (or origination) rate per unit of time (Van Valen 1984; Foote 2000). To distinguish between origination and immigration during faunal turnover, we used the PBDB to check for the presence of “new” genera in older marine

biodiversity hotspots (i.e., the West Tethyan and/or Arabian).

Overall differences in the presence/absence of genera in communities from different time periods were expressed as a Bray–Curtis dissimilarity matrix (Bray and Curtis 1957), which we compared with a corresponding matrix of the time differences between each pair of communities, using a Mantel test (Mantel 1967) for correlation between matrices based on random permutations of the dependent (Bray–Curtis) matrix. Each data point thus represents a comparison between a pair of communities, relating the similarity in their composition to the amount of time separating them. The degree of change in community composition over time is expressed in bivariate plots that relate taxonomic dissimilarity to time separating them. Additionally, a hierarchical cluster analysis was performed on the Bray–Curtis dissimilarity matrix, showing the distinctions in taxonomic composition of coral communities. Statistical analyses on taxonomic turnover were carried out using R, Version 2.15.2 software (R Development Core Team 2012).

Functional Traits.—To investigate functional diversity of corals in the Central Indo-Pacific, we collected information on five morphological traits: colony attachment, corallum type, colony growth form, corallite arrangement, and corallite size. The morphological characters were selected because they reflect coral functional strategies (Rachello-Dolmen and Cleary 2007; Darling et al. 2012; Sommer et al. 2014). For example, branching colonies are usually fast growing and therefore more successful than other forms at occupying space and acquiring light and thus at obtaining key resources (Chappell 1980; Baird and Hughes 2000). However, they are sensitive to high hydrodynamic energy and thermal stress (Chappell 1980; Madin 2005; McClanahan et al. 2007). In contrast, massive colonies are resistant to high hydrodynamic energy and high sedimentation (Chappell 1980; Jackson and Hughes 1985; Soong 1993; Rachello-Dolmen and Cleary 2007); therefore, they are more common than other colony morphologies in marginal reef environments (Sommer et al. 2014).

All five morphological characters were assessed using both our fossil coral collection

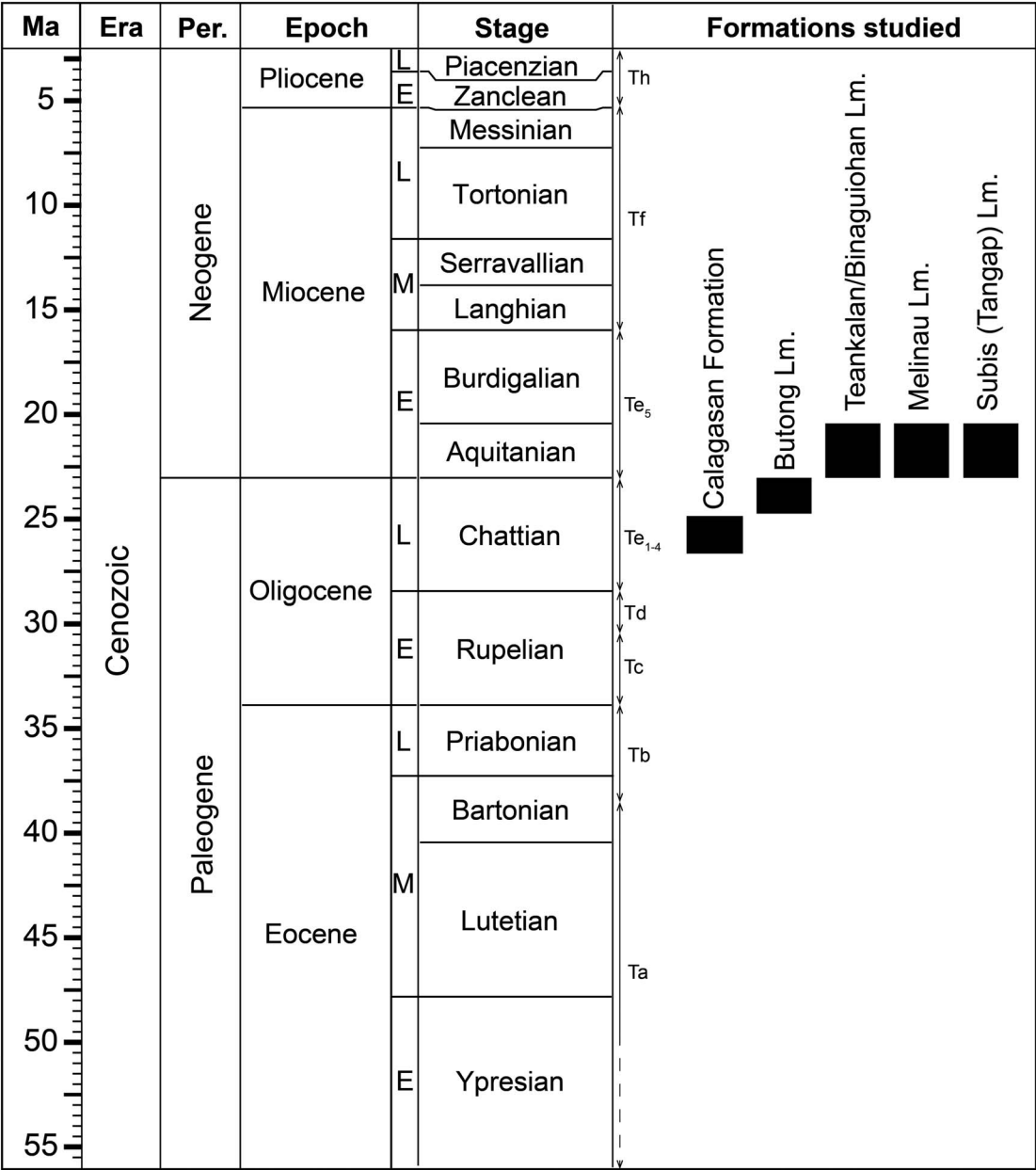


FIGURE 2. Study interval with temporal range of the formations studied. E, early; M, middle; L, late. Large benthic foraminifera zones indicated (Ta–Th).

and photographs from the taxonomic literature. For the colony attachment trait, corals were defined as either *attached* or *free-living*. For the corallum type trait, corals were assigned to either *colonial* or *solitary*. The colony growth form trait was categorized into *branching*, *discoïd*, *platy*, *massive*, and *cup-shaped*. The

branching category included all corals with branching and columnar growth forms. The *platy* category refers to vertical, horizontal, unifacial, or bifacial thin tiers. Encrusting colony growth forms were included in the *massive* category because preservation in coral fragments can make it difficult to differentiate

thick encrusting sheets from small massive colonies. A similar grouping has been used to characterize coral growth forms in modern reefs (Darling et al. 2012; Sommer et al. 2014). The corallite size trait was classified into five categories by averaging the measurements of corallite diameter/valley width of all newly collected specimens within a genus (three corallite measurements per specimen): *very small* (<3 mm), *small* (3–6 mm), *medium* (6–8 mm), *large* (8–15 mm), and *very large* (>15 mm). The corallite arrangement trait referred to the relative position of the corallites within the colony: *cerioid*, *plocoid*, *thamasterioid*, *meandroid*, *phaceloid*, and *single*. All functional character states were treated as nested, binary characters (present/absent), because some genera can exhibit multiple character states for the same functional trait (e.g., the same genus can have both small and medium corallite sizes).

Functional Diversity.—Functional diversity of Central Indo-Pacific corals was estimated using Rao's diversity coefficient (also known as quadratic entropy) (Rao 1982), which measures the mean functional distances between randomly selected genus pairs based on the abundance-weighted variance of the dissimilarities computed among all pairs of genera (Rao 1982; Champely and Chessel 2002; Ricotta 2005). Rao's diversity coefficient has been shown to be a good estimate of functional diversity (Botta-Dukát 2005; Scherer-Lorenzen et al. 2007; Weigelt et al. 2008). For functional diversity analysis, we used the mixed-variables coefficient of distance, as our functional trait states are independent or nested binary variables (Pavoine et al. 2009). The mixed-variables coefficient of distance generalizes Gower's coefficient, maintaining the Euclidian properties (Gower and Legendre 1986; Champely and Chessel 2002; Pavoine et al. 2009; Laliberté and Legendre 2010). All calculations were completed using the 'ade4' package in R (Pavoine et al. 2009).

Functional Turnover.—Origination and extinction rates were calculated for each functional character state through time. To explore how diversity of functional groups varied through time, taxonomic diversity analyses were repeated separately for each individual functional character state. For

example, taxonomic diversity was calculated using only the coral taxa with a massive colony growth form. Overall differences in the relative abundance of functional character states between each pair of studied time intervals were calculated (explained above in "Taxonomic Turnover") using a Bray-Curtis dissimilarity index (Bray and Curtis 1957) and visualized using a nonmetric multidimensional scaling (nMDS) ordination (Anderson 1971; Giraudel and Lek 2001). Relative abundance of a functional character state was derived by dividing the absolute number of genera with that functional trait state (e.g., branching) in particular time interval by the total number of genera present in that time interval. Rotational vector fitting was used to relate the functional character states to the ordination of time intervals, allowing us to quantify the strength of this relationship through a correlation coefficient (r^2) (Faith and Norris 1989; King and Richardson 2008). The vectors point in the direction of the gradient, with the length of the vector indicating the strength of the gradient (i.e., the correlation between the ordination and the functional character states). Significance of vectors was estimated using 999 random permutations (Oksanen et al. 2013). As with the taxonomic data, a Mantel test and a hierarchical cluster analysis were performed on the Bray-Curtis dissimilarity matrix based on the abundances of the functional trait states. Statistical analyses on functional turnover were carried out using the 'vegan' (Oksanen et al. 2013) and 'hclust' (Murtagh 2000) packages in R.

Environmental Drivers.—nMDS ordination of the Bray-Curtis dissimilarity matrix was used to visualize the relative differences in taxonomic and functional trait composition among time intervals (Anderson 1971; Giraudel and Lek 2001). To identify the main environmental drivers of taxonomic diversity and abundance of functional traits through time, we used rotational vector fitting of environmental factors (Faith and Norris 1989; King and Richardson 2008) onto the nMDS ordination. We used global sea level, Northern Hemisphere temperature, global deep-water temperature (de Boer et al. 2012), and global benthic $\delta^{13}\text{C}$ (proxy for productivity) (Zachos et al. 2001). These factors were selected because

of their significance to development of reef environments (Wilson 2008) and availability of their continuous, long temporal records. Although coral communities are influenced by both global and local environmental factors, records of local factors such as salinity, riverine discharge, turbidity, or upwelling lack a long temporal record. Significance of each environmental factor was determined using 999 permutations. To capture fluctuations in the environmental factors within each time interval, the minimum, maximum, mean, and variance (based on standard deviation) of each factor over the entire study interval were used as separate variables. Prior to the nMDS and rotational vector analyses, we checked for multicollinearity of environmental variables using Spearman's correlation coefficient with a cutoff of $\rho > 0.8$. After excluding correlated environmental factors, we tested the correlation of five global environmental factors (sea-level mean, sea-level variance, deep water–temperature variance, and productivity mean and variance) with coral community composition.

Results

Biostratigraphy.—The co-occurrence of the foraminifera *Eulepidina*, *Vlerkina borneensis*, and *Tansinhokella* at the Cebu localities was indicative of the Te_{2-4} LBF zone (Oligocene: middle Chattian, ~26.5 Ma). In the Calagasan Formation, *V. borneensis* was abundant and *Tansinhokella* was rare, whereas in the Butong Limestone Formation, *Tansinhokella* was more common than in the Calagasan Formation and *Austrotrillina* was also present. This variation between the two formations indicates that the Calagasan Formation is slightly older (Oligocene: middle Chattian) than the Butong Limestone Formation (Oligocene: late Chattian, ~24.5 Ma). Due to the presence of *Nephrolepidina*, *Eulepidina*, *Miogypsina*, and *Miogypsinoides*, the Negros locality dates to the Te_5 LBF zone (early Miocene: Aquitanian, ~23 Ma) as do the localities from Sarawak. Therefore, sites from Cebu are Chattian (~28.4–23 Ma) in age, whereas the Sarawak and Negros sites are from the Aquitanian (~23–20.4 Ma) (Fig. 2).

Sampling Adequacy and Estimated Stratigraphic Ranges.—We identified a total of 30 genera

from the 1606 coral specimens that we collected (Fig. 3). Cebu has the largest number of genera (27 in 1489 samples), Sarawak the second largest (14 in 106 samples), and Negros the fewest (3 in 10 samples) (Table 2). The sampling curve for the Chattian begins to level off after 500 specimens. The Aquitanian curve follows the same trajectory but does not reach an asymptote, indicating undersampling (Fig. 4) (for more information on sampling adequacy and bias, see Supplementary Material Appendix 1). Nevertheless, our new fossil collection extends the FAD in the Central Indo-Pacific for *Acanthastrea*, *Astrea*, *Coelastrea*, and *Lobophyllia* into the Chattian and for *Blastomussa* into the Aquitanian. Refer to Supplementary Figure 1 for a full listing of estimated temporal ranges for all fossil corals used in our diversity analyses and to calculate taxonomic and functional diversities in the Central Indo-Pacific.

Taxonomic Diversity.—A continuous but gradual increase in MSD of Central Indo-Pacific fossil coral genera occurred throughout the whole study interval (Fig. 5A). The most rapid increases in generic diversity (MSD) occurred between the Rupelian and Chattian with 21 new genera and between the Chattian and the Aquitanian with 20 new genera. In the Chattian, 29% of 21 newly occurring genera arose through speciation and 71% migrated mostly from older marine diversity hotspots (i.e., the West Tethyan and/or Arabian). The Aquitanian shows a very similar pattern (30% speciation, 70% migration) (Supplementary Table 3). The Chao2 diversity curve mirrors MSD, although diversity estimates range from 5 to 40% higher depending on the time interval. The overall increasing pattern in generic diversity shown by Chao2 shows two peaks: one in the Tortonian and a higher one in the Zanclean.

We found a positive logarithmic relationship (for MSD: $r^2 = 0.87$, $p < 0.05$) between coral diversity (MSD or Chao2) and the relative area of coral-dominated carbonates, with a distinct pattern of increased diversity over time (Fig. 6). However, patterns are visible when estimating diversity from individual time epochs: (1) from the early Eocene to the early Oligocene, coral diversity gradually increased, but the area of

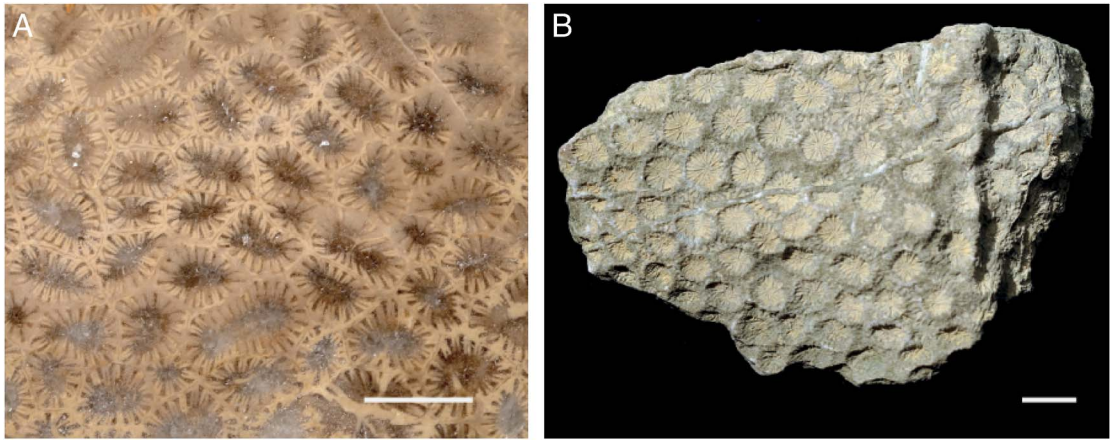


FIGURE 3. Representative example of corals collected from (A) limestone outcrops, *Hydnophora* sp., QMF58230, and (B) shale outcrops, *Astrea* sp. QMF 58494. Scale bars, 1 cm.

TABLE 2. Number of coral genera relative to the number of collected specimens found at each of the three study sites and in the two investigated time stages.

	Number of genera	Number of coral specimens
Calagasan Formation	22	915
Butong Limestone Formation	20	573
Teankalan/Binaguiohan Limestone	4	11
Melinau Limestone	6	33
Subis (Tangap) Limestone	10	73
Middle Chattian	22	915
Late Chattian	21	574
Aquitanian	14	116

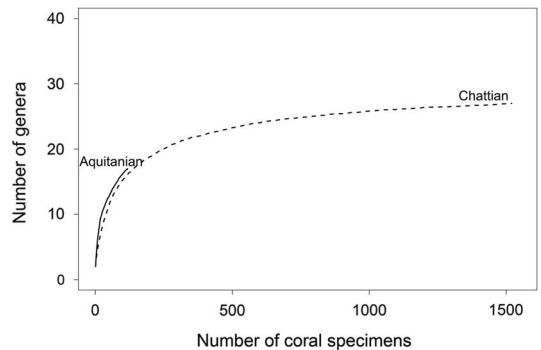


FIGURE 4. Coral genera sampling curve for all Chattian (dashed line) and Aquitanian (solid line) specimens collected from Malaysia (Sarawak) and the Philippines (Negros and Cebu).

coral-dominated carbonates does not increase proportionally (as fast); (2) diversity increases more rapidly in the late Oligocene and the early Miocene, when there is an accompanying abrupt increase in the area of coral-dominated carbonates; and (3) a gradual increase in coral diversity continues into the Pliocene despite the contraction in area of coral-dominated carbonates.

Taxonomic Turnover.—Several origination/immigration and extinction/emigration periods were identified (Fig. 7). Origination rates were relatively high during most of the Eocene, even at the end, when an extinction event (seven genera within ~3.3 Ma) took place. From the Oligocene to the early Miocene, origination rates increased, culminating in the highest

recorded origination rate (19 new genera within ~2.6 Ma) in the Aquitanian, when the extinction of six genera also occurred. For most of the rest of the Miocene, origination and extinction rates were relatively low yet remained constant, but in the Messinian a high extinction rate (nine genera within ~1.9 Ma) occurred. In the Pliocene, a high origination rate in the Zanclean and a high extinction rate in the Piacenzian were recorded.

Dissimilarity in generic composition correlated significantly with the amount of time separating time intervals, that is, the difference in generic composition increases as the time difference between two communities increases (polynomial model $r^2=0.85$, $p<0.05$; Mantel

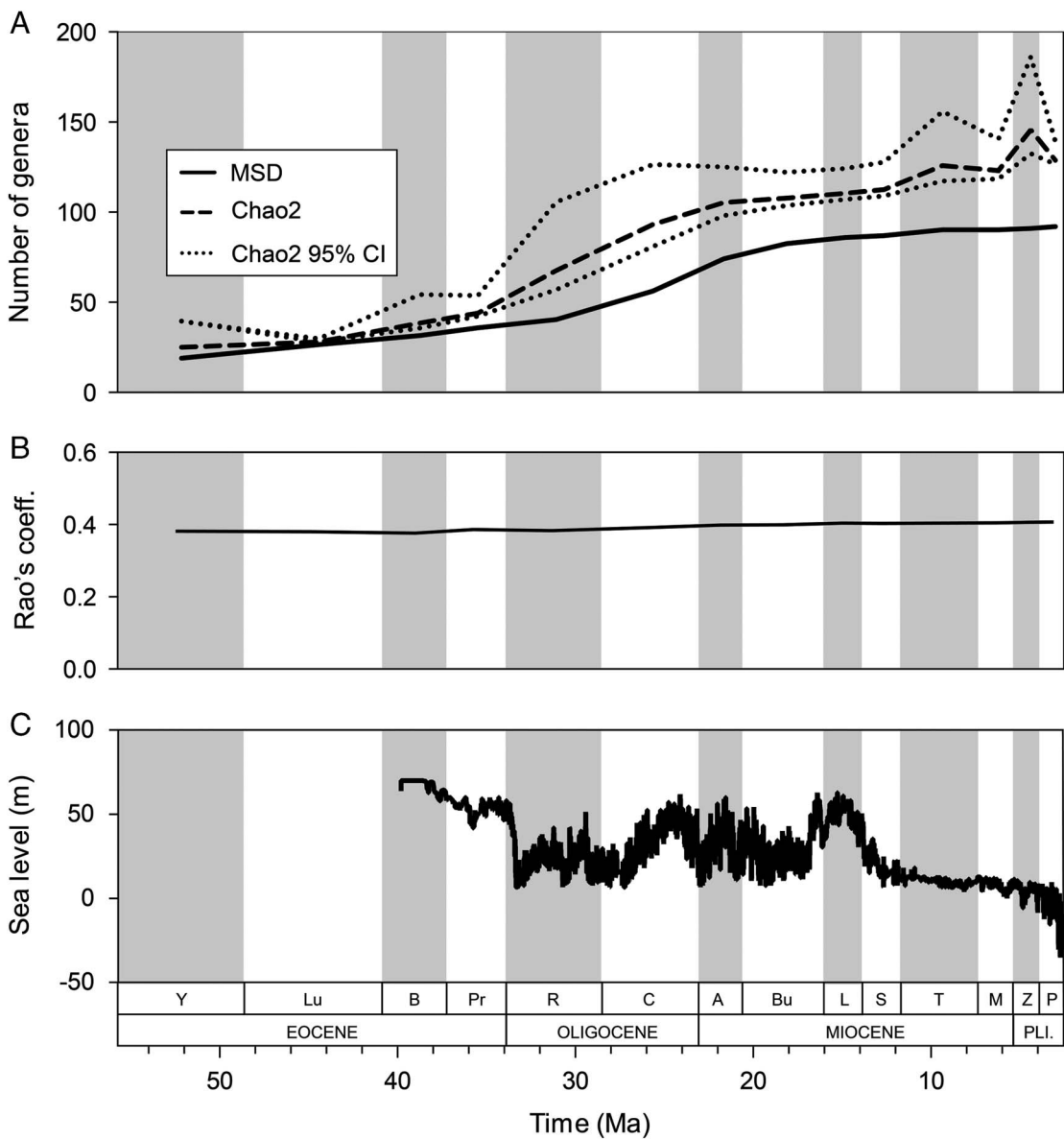


FIGURE 5. Coral diversity in the Central Indo-Pacific and global sea level through time. A, Taxonomic diversity estimated using two methods: the MSD and the Chao2 index with lower and upper confidence intervals; B, functional diversity calculated using Rao's coefficient; C, global sea level (de Boer et al. 2012). CI, confidence interval; MSD, mean standing diversity.

test $r=0.91$, $p=0.0001$) (Fig. 8A). Hierarchical cluster analysis (Fig. 8B) shows that Rupelian generic composition is most similar to the composition of older Eocene communities, whereas the generic composition of Chattian coral communities is most similar to those from the younger Miocene and Pliocene time periods. Moreover, overall generic composition

becomes more similar over time. For instance, there is greater similarity between the Miocene and Pliocene assemblages (~ 0.2 dissimilarity) than there is between earliest Miocene (Aquitainian) and late Oligocene (Chattian) assemblages (~ 0.3 dissimilarity) or early Oligocene (Rupelian) and Eocene assemblages (~ 0.7 dissimilarity).

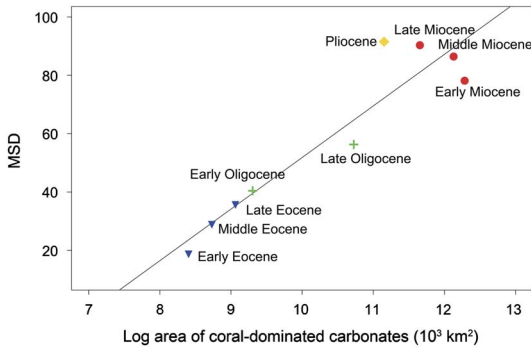


FIGURE 6. Plot of the area of coral-dominated carbonates (log) (from Wilson 2008) against the MSD of coral genera from the early Eocene to the Pliocene of the Central Indo-Pacific.

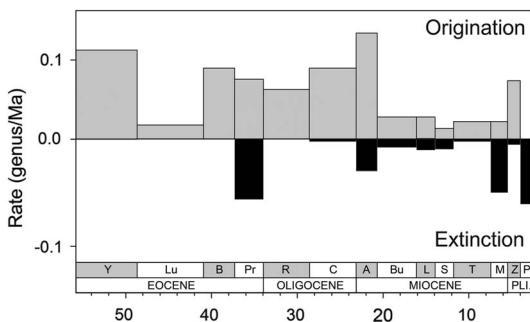


FIGURE 7. Origination and extinction rates of Central Indo-Pacific coral genera through time (data from our new collection combined with records of fossil corals derived from the PBDB). Geologic stage abbreviations: Y, Ypresian (~55.8–48.6 Ma); Lu, Lutetian (~48.6–40.4 Ma); B, Bartonian (~40.4–37.2 Ma); Pr, Priabonian (~37.2–33.9 Ma); R, Rupelian (~33.9–28.4 Ma); C, Chattian (~28.4–23 Ma); A, Aquitainian (~23–20.4 Ma); Bu, Burdigalian (~20.4–15.97 Ma); L, Langhian (~15.97–13.82 Ma); S, Serravallian (~13.82–11.6 Ma); T, Tortonian (~11.6–7.2 Ma); M, Messinian (~7.2–5.3 Ma); Z, Zanclean (~5.3–3.6 Ma); P, Piacenzian (~3.6–2.588 Ma); PLI, Pliocene.

Functional Diversity.—All functional character states are present throughout the study interval, with the exception of phaceloid corallite arrangement in the Ypresian and the Lutetian. The functional diversity of corals (Rao's diversity coefficient) from the Central Indo-Pacific is relatively constant across the entire time interval, yielding an average value of 0.39 ± 0.02 (Fig. 5B).

Generally, when comparing the functional character state abundances over time, the time intervals that are closest together tend to

be the most similar in their functional composition (polynomial model: $r^2 = 0.84$, $p < 0.05$; Mantel test: $r = 0.89$, $p = 0.0001$) (Fig. 8C). Hierarchical cluster analysis shows that, functionally, the Eocene and Oligocene coral communities are more similar to each other than they are to Miocene and Pliocene ones (Fig. 8D), which is in contrast with taxonomic composition, where the taxonomic composition of the Chattian (late Oligocene) is more similar to the younger assemblages than the older (Fig. 8B).

Functional character trait-state abundances vary through time, with many trait states correlating with specific time intervals (Fig. 9). The nMDS ordination of functional trait abundances (Fig. 9) corroborates the results of the Mantel test and hierarchical cluster analysis (Fig. 8C,D) with coral communities from time intervals closer in age plotting closer to each other, that is, characterized by similar trait states. For instance, difference in functional character states in the Aquitainian and Burdigalian, when compared with other time intervals, are driven primarily by an increased occurrence of corals with platy (P) colony morphology, meandroid corallite arrangement (me), and large (l) and very large (vl) corallite sizes.

Although there are distinct generic origination/immigration and extinction/emigration peaks, trait originations/immigrations and extinctions/emigrations occur throughout the study interval, and origination/extinction rates are unique for each functional character state through time (Supplementary Fig. 2). Origination rates vary among functional character states, but all increase sometime during the initiation and strong pulse of origination/immigration between the Rupelian and Aquitainian (Supplementary Fig. 2). During periods with high extinction rates (Priabonian, Aquitainian, Messinian, and Piacenzian), extinctions occur across all functional character states except free-living, branching, platy, cerioid, and phaceloid in the Priabonian; free-living, cup-shaped, and cerioid in the Aquitainian; platy, cerioid, phaceloid, and thamnasterioid in the Messinian; and branching, cup-shaped, cerioid, plocoid, phaceloid, and very small and medium-sized corallites in the Piacenzian.

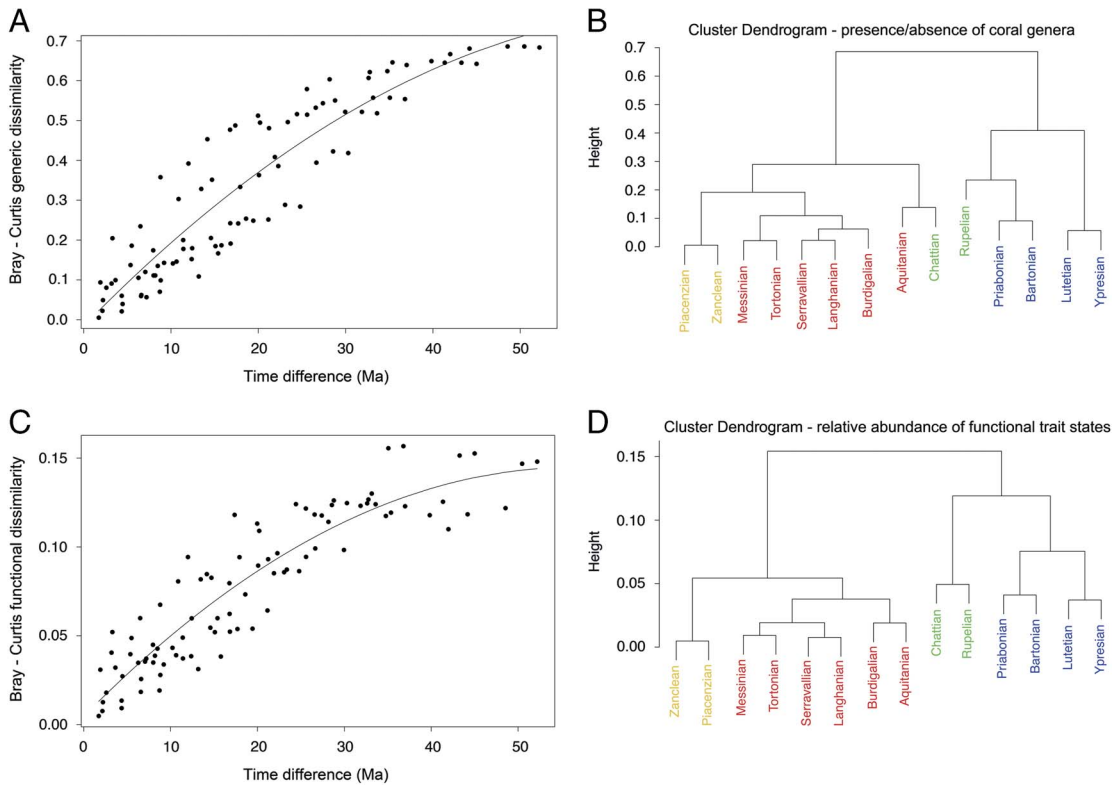


FIGURE 8. A, Bray-Curtis dissimilarity of coral community composition (genus level) plotted against time separating each pair of coral assemblages; B, cluster dendrogram for coral communities (genus level) in each time interval; C, Bray-Curtis dissimilarity of functional trait-state abundances plotted against time separating each pair of coral assemblages; and D, cluster dendrogram for functional trait-state abundances. Time intervals: blue, Eocene; green, Oligocene; red, Miocene; yellow, Pliocene.

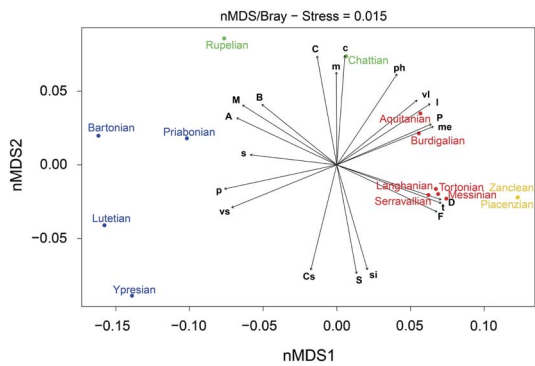


FIGURE 9. nMDS ordination of abundance of functional trait states through time. Rotational vectors for functional trait-state abbreviations: A, attached; F, free-living; C, colonial; S, solitary; B, branching; M, massive; P, platy; D, discoid; Cs, cup-shaped; c, cerioid; p, plocoid; t, thamnasterioid; me, meandroid; ph, phaceloid; si, single; vs, very small (<3 mm); s, small (3–6 mm); m, medium (6–8 mm); l, large (8–15 mm); vl, very large (>15 mm). Time intervals: blue, Eocene; green, Oligocene; red, Miocene; yellow, Pliocene.

Environmental Drivers.—Community composition based on the ordination of the presence/absence of coral genera varies through time, specifically with communities from the Eocene (Ypresian–Priabonian) and early Oligocene (Rupelian) plotting separately from younger time intervals in the nMDS ordination (Fig. 10A), corroborating the results of the hierarchical cluster analysis. Overall differences of taxonomic community composition appear to be associated with global sea-level mean, given that it is the only significant environmental factor ($r^2=0.67$, $p<0.05$). Functional community composition shows a similar division between older (Eocene, Oligocene) and younger (Miocene, Pliocene) assemblages, with the Eocene more associated with sea-level mean, the Oligocene with sea-level variance, and the Miocene with both global sea-level mean ($r^2=0.58$, $p<0.05$) and sea-level variance ($r^2=0.58$, $p<0.05$) (Fig. 10B).

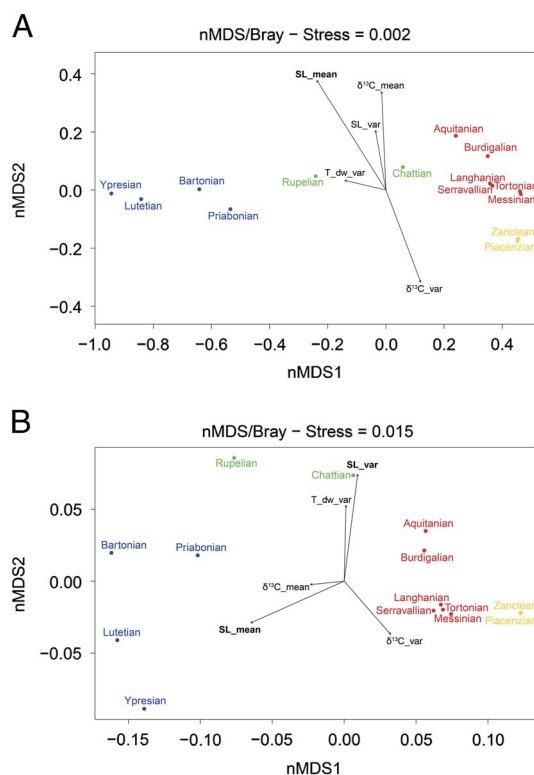


FIGURE 10. Similarity among time periods for community composition in relation to environmental variables. nMDS ordination of time periods as a function of the presence/absence of coral genera (A) and as a function of relative abundance of functional trait states (B). Time intervals: blue, Eocene; green, Oligocene; red, Miocene; yellow, Pliocene. Rotational vectors for environmental variables are fitted to the ordination axis to reveal their corresponding relationship to time intervals; significant environmental factors are highlighted in bold ($p = 0.05$).

Discussion

Biodiversity hotspots are regions with sharply higher diversity relative to surrounding regions. They vary temporally and spatially due to the interplay of modern and historical ecological, geological, and environmental factors. Biodiversity itself is not only a measure of taxonomic richness but also of organismal and ecosystem functionality. Here we couple the taxonomic and functional development of coral communities with environmental dynamics of biodiversity hotspots to understand what drives biogeographic patterns of biodiversity and species–area relationships over time. Our findings show the Central Indo-Pacific biodiversity hotspot originated during

the Oligocene. This provides a new working hypothesis for the timing of origination of the present-day Indo-Pacific diversity hotspot, clarifies previous estimates of the timing of origination (Wilson and Rosen 1998; Renema et al. 2008; McMonagle 2012; Santodomingo et al. 2016) and gives insight into underlying mechanisms of hotspot development. Specifically, our data show variable rates in the accumulation of diversity through time and decoupling of taxonomic and functional diversity patterns.

Taxonomic Diversity.—Low diversity in the Eocene followed by a rapid increase in the Oligocene and earliest Miocene (Rupelian to Aquitanian, Figs. 5A and 7) and the maintenance of relatively high diversity throughout the rest of the Miocene and Pliocene results from the interactions between origination and extinction rates (Fig. 7). It should be noted that the increase in diversity might be underestimated due to sampling of indurated limestone lithologies, which represent potentially the most diverse depositional environments in the Central Indo-Pacific (Wilson 2015). Limestones are challenging for studying macrofossils due to diagenetic alteration (Wright and Burgess 2005), and this process is suggested by the undersampling of the Aquitanian corals in our new fossil collection (Fig. 4). As such, more complete sampling of limestone lithologies would enhance the observed increase in taxonomic diversity.

The complex interplay among, and co-occurrence of, global and local tectonic, eustatic, oceanographic, and climatic events (TECO events of Rosen 1984) makes it challenging to determine the precise drivers of the observed patterns in origination and extinction as well as coral community composition (Wilson 2008). Nonetheless, increasing taxonomic diversity from the Eocene to Pliocene (Fig. 5A) appears to be negatively (Fig. 5C) associated with mean global sea level (Fig. 10A). An overall decrease in global sea level from the Eocene to the Oligocene (Fig. 5C) coincides with local tectonics, such as continental collision and regional uplift (Hall 2002; Hutchison 2004), that resulted in an increase in shallow-marine habitats. It is possible that a relatively high sea level largely constrained

coral proliferation and diversification in the Eocene (Fig. 5C). We propose that falling sea level from the Eocene until the late Oligocene largely promoted coral proliferation and diversification by increasing the area of shallow-water habitats ideal for coral growth (Fig. 11) (Hall 2001; Renema et al. 2008; Wilson 2008). Extinction at the end of the Eocene likely reflects an alternative response of some taxa to the environmental changes associated with abrupt sea-level fall; however, the loss of these taxa is not enough to negate the general increasing trend in taxonomic diversity over time (Fig. 5A).

During the late Oligocene–early Miocene, the greatest increase in taxonomic diversity occurs (Fig. 5A), likely representing the accumulation of taxonomic diversity due to sustained, relatively higher origination rates from the Bartonian to the Aquitanian (Fig. 7). This increase in taxonomic diversity coincides with the abrupt regional increase in the relative area of coral-dominated carbonates (Wilson 2008), resulting from the transition of local reef habitats from large carbonate platforms dominated by foraminifera in the Eocene to more isolated and shallower platforms dominated by corals in the Miocene (e.g., Wilson 2002; Mihaljević et al. 2014). Observation of coral diversity peaking in the Aquitanian, when the relative area of coral-dominated carbonates abruptly increased (Wilson 2008), is consistent with a species–area relationship, in which the number of species is expected to increase as the amount of available habitat space increases (Rosenzweig 1995). Lack of reef framework in the Central Indo-Pacific until the Aquitanian (Wilson 2002, 2008; Mihaljević et al. 2014) suggests that the Central Indo-Pacific diversity hotspot originated (i.e., increased faunal turnover and magnitude of diversity) in habitats characterized by low abundance but moderate diversity of corals (Mihaljević et al. 2014). The positive logarithmic trend between the MSD and area of carbonate structures (Fig. 6) supports this hypothesis.

Interestingly, there is a lack of a species–area relationship from the early Miocene onward. Coral diversity continued to increase through the Pliocene, despite a decrease in origination rates from the early Miocene and decreasing

area of coral-dominated carbonates from the middle Miocene (Fig. 6). If the area of coral-dominated carbonates is considered a proxy for suitable habitat, it is plausible that as habitat availability declined from the early Miocene, physical barriers could have developed, creating isolation and promoting speciation (i.e., continued diversification) (Keith et al. 2013). An increase in habitat complexity could sustain a continued, albeit relatively lower, rate of diversification into the Pliocene (Guégan et al. 1998; Báldi 2008). Additionally, if habitat area was reduced and coral taxa were confined to coexist in a smaller area, increased pressure from biological interactions (e.g., competition) also could have contributed to low diversification rates (e.g., Rosenzweig 1995) and the lack of a species–area relationship through to the Pliocene.

The decrease in shallow-marine habitats from the middle Miocene is likely related to the complex interactions between continued falling sea level and regional tectonic uplift. From the Eocene to the Oligocene, the interaction of falling global sea level and regional uplift resulted in the expansion of shallow-marine habitats by collectively reducing regional sea level. By the middle Miocene, however, the effects of this interaction appear to have changed, as the trajectory between global sea level and the area of coral-dominated carbonates reverses (Fig. 11). We propose that, through the interplay of falling global sea level and regional tectonic uplift, regional sea level reached a critical (low) threshold in the Central Indo-Pacific around the middle Miocene, after which continued sea-level fall (and uplift) began to decrease the area of shallow-marine habitats suitable for corals. This transition exemplifies the complexity of sea-level and tectonic interactions and their cumulative effect on ecology over time.

In addition to the interaction of global sea level and local tectonics, other global and local environmental variables, with less continuous records than sea level, could have influenced coral diversity trends over time. For instance, changes in ocean chemistry might have promoted the abrupt increase in coral diversity from the Rupelian to the Aquitanian (across the Oligocene/Miocene boundary). During this

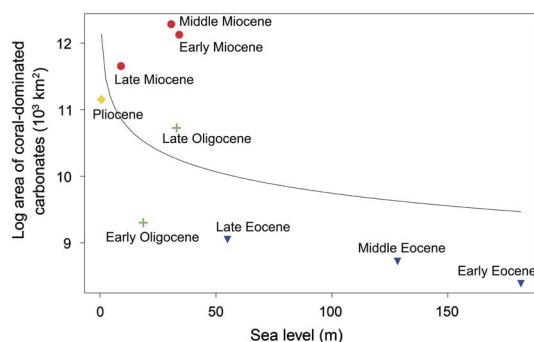


FIGURE 11. Plot of the area of coral-dominated carbonates (log) and sea level from the early Eocene to the Pliocene of the Central Indo-Pacific.

time, the global seawater Mg/Ca ratio (a proxy for carbonate precipitation for which higher values favor the precipitation of aragonite over calcite) was increasing, and local salinity became more consistent due to more constant terrestrial runoff driven by a climatic shift—from seasonal to ever-wet conditions—over the Oligocene/Miocene boundary (Stanley and Hardie 1998; Morley 2000, 2011; Morley et al. 2003). The shift to more consistent, less seasonably variable conditions would lead to more favorable, less stressful shallow-water environmental conditions, potentially promoting coral proliferation and diversification (Wilson 2008).

Johnson et al. (2015) highlighted that imprecise dating methods in early Central Indo-Pacific collections led to some genera being placed in incorrect time periods. Considering these results, the high extinction and origination rates observed in our data at the end of the Miocene and beginning of the Pliocene, respectively, should both be lower. Consequently, lower extinction rates would correspond to relatively higher diversity and limited turnover throughout the Miocene and Pliocene, corroborating findings of previous studies (Johnson et al. 2015; Santodomingo et al. 2016). Although these findings would slightly alter magnitude of the observed diversity patterns in the latest Miocene and early Pliocene, they do not affect the estimated timing of origination of the Central Indo-Pacific biodiversity hotspot, which our results show to be in the late Oligocene.

Functional Diversity.—Intriguingly, observed patterns in taxonomic diversity are decoupled from patterns in functional diversity. Despite significant fluctuations in taxonomic diversity, functional diversity remained relatively stable (Fig. 5B), irrespective of temporal variations in trait abundances and turnover rates (Fig. 9 and Supplementary Fig. 2). Taxonomic and functional diversity trends could be expected to be similar if new traits are arising as the result of the development of new taxa (or vice versa) (Naeem and Wright 2003; Petchey and Gaston 2006; Villéger et al. 2010). Our finding of high stability in functional diversity during periods of high diversification contradicts these expectations and points toward high functional redundancy, suggesting that species added to the community occupy the same functional space (i.e., have similar traits) as already existing species (Mayfield et al. 2010; Cadotte et al. 2011).

The co-occurrence of species occupying the same functional space can be facilitated through expansion of habitat area, resulting in relaxation of species competition (e.g., space). As previously discussed, global sea-level changes combined with local tectonics contributed to the expansion of shallow-marine habitats suitable for coral growth. The influence of the sea level on abundance of functional trait states is clearly illustrated by the separation of Eocene–Oligocene and Miocene–Pliocene functional communities by significant sea-level mean and variance vectors (Fig. 10B). Additionally, the reported increase in habitat complexity (Mihaljević et al. 2014) might further promote functional redundancy in corals. While most functional trait states were present in all time intervals, that is, functional diversity through time is constant, their relative abundances varied through time in response to environmental changes. Considering high morphological plasticity of corals, simple turnover of already existing trait states would allow for rapid adaptation to the emerging habitat availability and complexity without the “need” for new adaptations. Thus, functional redundancy might have contributed to the ~28 Myr-long persistence of the Central Indo-Pacific biodiversity hotspot through promoting ecosystem stability following declines in

TABLE 3. Percentage contribution of old and new taxa to relative abundances of coral growth forms. Old taxa refers to 46 genera already present in Rupelian.

	Rupelian		Chattian		Aquitanian	
	Old (%)	New (%)	Old (%)	New (%)	Old (%)	New (%)
Branching	100	0	77.27	22.73	60.71	39.29
Massive	100	0	69.77	30.23	58.82	41.18
Platy	100	0	52.94	47.06	42.86	57.14
Discoid	100	0	57.14	42.86	36.36	63.64
Cup-shaped	100	0	80.00	20.00	57.14	42.86

taxonomic diversity (Fonseca and Ganade 2001; Guillemot et al. 2011).

Despite the decoupling of taxonomic and functional trait diversity patterns, both taxonomic and functional trait composition become more dissimilar as the time separating assemblages increases (Fig. 8A,C). This ubiquitous increase in dissimilarity of assemblages with increasing time is likely not solely driven by random accumulation of diversity over time, but might also reflect the development of reef communities in response to habitat availability, for example, diversity increases with increasing habitat availability (Tager et al. 2010). Temporal trends in taxonomic, trait, and environmental conditions are not linear per se (i.e., taxonomic diversity has not increased at a steady rate over time, nor has sea level linearly lowered), and fluctuations in these factors likely have interactive effects on coral assemblages over time (Bromfield and Pandolfi 2011), resulting in dissimilar communities from the Eocene to the Pliocene (Fig. 8B,D).

Although cluster analyses show increasing dissimilarity with time for both taxonomic and functional trait assemblages, the results also reveal that the pattern of dissimilarity is not the same for both taxonomic and functional assemblages (Fig. 8B,D). Most notably, Chattian generic composition was more similar to that of younger time periods (Miocene and Pliocene) than to the Eocene, whereas the Chattian functional composition was more similar to the older (Eocene) period (Fig. 8B,D). This implies that the biggest taxonomic turnover (Rupelian–Chattian) is largely disconnected from functional trait dynamics. The decoupling of Chattian taxonomic and functional assemblages suggests that the response of corals to expanding habitats in the Chattian was characterized by the local appearance of

new taxa (leading to the grouping of this era with the younger time periods) that maintained the same functional space as those in the pre-Chattian (therefore maintaining functional similarity to older time periods). In contrast, further expansion and diversification of suitable habitats in the Aquitanian led to the emergence of new taxa that was accompanied by changes in the functional space of the Aquitanian coral communities (Renema et al. 2008; Wilson 2008; Mihaljević et al. 2014). The decoupling of taxonomic and functional diversity means that (1) functional trait dynamics are not dependent on taxonomic dynamics (Figs. 5 and 8), and (2) taxonomic and trait dynamics can be influenced differently by the same environmental factors (Fig. 10).

Mechanisms of Increase in Coral Reef Area during the Origin of the Hotspot.—The abrupt increase in reef area in the late Oligocene and especially the early Miocene is associated with the proliferation of already present taxa and origination/immigration of new taxa. The strong positive correlation between coral-dominated area and MSD indicates that the origination/immigration of new taxa was undoubtedly a key mechanism. However, the proliferation of already present taxa must also have contributed to the abrupt increase in coral-dominated carbonates because: (1) increases in the area of coral-dominated carbonates and coral diversity are not proportional through time, and (2) Chattian communities are functionally more similar to communities older than the Miocene. For example, a comparison of the contribution of already present and new taxa to the relative abundance of colony growth trait states (branching, massive, platy, discoid, and cup-shaped)—a proxy for coral area—revealed that taxa already present in the

Chattian contributed more to the relative abundance of all coral growth forms than did new taxa. The same is true in the Aquitanian, with the exception that new taxa with platy and discoid growth forms, which tend to be common in turbid (oligophotic) environments (Rosen et al. 2002; Browne et al. 2012; Novak et al. 2013), contribute more than already present taxa with these growth forms (Table 3). These patterns suggest that the Central Indo-Pacific biodiversity hotspot has developed in oligophotic conditions in two phases: (1) in the Chattian through proliferation of coral genera already present since the Rupelian, and (2) in the Aquitanian through the expansion of existing taxa as well as through the origination/immigration of new genera.

Conclusions.—Our study shows that the Central Indo-Pacific has persisted as a biodiversity hotspot since the Chattian (~28 Myr), which is ~5 Myr earlier than previously thought. We postulate that functional redundancy, seen through the decoupling of taxonomic and functional diversity, potentially contributed to the persistence of the hotspot by promoting ecosystem stability. An increase in shallow-marine habitats suitable for coral growth, resulting from the interplay of global sea level and regional tectonics, seems to have driven this diversification of Central Indo-Pacific corals, which, through proliferation of already present genera, immigration, and origination of new ones contributed to the expansion of reef area. Therefore, the history of the Central Indo-Pacific highlights that marine biodiversity hotspots develop through a complex interplay of proliferation, accumulation, and origination of coral genera.

Acknowledgments

We acknowledge financial contributions from Petrolam Nasional Berhad (Petronas), Pearl Energy, the University of Queensland (UQ) International Scholarship (UQI) 2011–2014, the UQ Collaboration and Industry Engagement Fund, UQ Centre for Marine Science, the Australian Research Council Centre of Excellence for Coral Reef Studies, and the Winifred Violet Scott Trust. We also thank the State Planning Unit in Kuching,

Malaysia for allowing us to conduct research in Sarawak, and the National Parks and Nature Reserves for a research permit for the Gunung Mulu National Park. Special thanks for logistical, physical, and all other help during seven field trips in Sarawak, Malaysia, and the islands of Cebu and Negros, Philippines, go to: Peter Lunt, the Gunung Mulu National Park staff (especially Brian Clark), Luke Southwell, Hollystone Quarry Sdn. Bhd. staff, people from the Long Jeh community, Brian Beck, King King Ting, the Mines and Geosciences Bureau in Manila and Mandaue City (especially Yolanda Aguilar and Abraham R. Lucero), Felix G. Nepomucedo, and Jack Coates-Marnane. We would also like to thank Simon Blomberg, Etienne Laliberte, Sandrine Pavoine, and Juan Carlos Ortiz for discussions about the statistical analysis. We thank Bas de Boer and James Zachos for access to raw environmental data. Special thanks to Aaron Hunter, Brigitte Sommer, Eugenia M. Sampayo, three anonymous reviewers and the editor for their comments on earlier versions of this article.

Literature Cited

- Adams, C. G. 1965. The Foraminifera and stratigraphy of the Melinau Limestone, Sarawak, and its importance in Tertiary correlation. *Quarterly Journal of the Geological Society* 121:283–338.
- Adams, C. G., and R. Haak. 1962. The stratigraphical succession in the Batu Gading area, Middle Baram, north Sarawak. The Geology and Mineral Resources of the Suai-Baram area, North Sarawak. Geological Survey Department (British Territories in Borneo) Memoir 13:141–150.
- Anderson, A. 1971. Ordination methods in ecology. *Journal of Ecology* 59:713–726.
- Aurelio, M. A., and R. E. Peña. 2010. Geology of the Philippines. Mines and Geosciences Bureau, Manila.
- Baird, A., and T. Hughes. 2000. Competitive dominance by tabular corals: an experimental analysis of recruitment and survival of understory assemblages. *Journal of Experimental Marine Biology and Ecology* 251:117–132.
- Báldi, A. 2008. Habitat heterogeneity overrides the species–area relationship. *Journal of Biogeography* 35:675–681.
- Barnes, E., M. A. Aurelio, C. Muller, M. Pubellier, R. D. Quebral, and C. Rangib. 1958. Geology and coal resources of the Argao-Dalaguete region, Cebu. Philippine Bureau of Mines, Manila.
- Bellwood, D. R., W. Renema, and B. R. Rosen. 2012. Biodiversity hotspots, evolution and coral reef biogeography: a review. Pp. 216–245 in D. J. Gower, K. G. Johnson, J. E. Richardson, B. R. Rosen, L. Rüber, and S. T. Williams, eds. *Biotic evolution and environmental change in Southeast Asia*. Cambridge University Press, New York.
- Botta-Dukát, Z. 2005. Rao's quadratic entropy as a measure of functional diversity based on multiple traits. *Journal of Vegetation Science* 16:533–540.
- Bray, J. R., and J. T. Curtis. 1957. An ordination of the upland forest communities of southern Wisconsin. *Ecological Monographs* 27:325–349.

- Bromfield, K. 2013. Neogene corals from the Indo-Pacific: Indonesia, Papua New Guinea, and Fiji. *Bulletins of American Paleontology* 387:1–136.
- Bromfield, K., and J. M. Pandolfi. 2011. Regional patterns of evolutionary turnover in Neogene coral reefs from the central Indo-West Pacific Ocean. *Evolutionary Ecology* 26:375–391.
- Browne, N. K., S. G. Smithers, and C. T. Perry. 2012. Coral reefs of the turbid inner-shelf of the Great Barrier Reef, Australia: an environmental and geomorphic perspective on their occurrence, composition and growth. *Earth-Science Reviews* 115:1–20.
- Budd, A. F. 2000. Diversity and extinction in the Cenozoic history of Caribbean reefs. *Coral Reefs* 19:25–35.
- Cadotte, M. W., K. Carscadden, and N. Mirotchnick. 2011. Beyond species: functional diversity and the maintenance of ecological processes and services. *Journal of Applied Ecology* 48:1079–1087.
- Champely, S., and D. Chessel. 2002. Measuring biological diversity using Euclidean metrics. *Environmental and Ecological Statistics* 9:167–177.
- Chao, A. 1984. Nonparametric estimation of the number of classes in a population. *Scandinavian Journal of Statistics* 11:265–270.
- Chao, A. A. 1987. Estimating the population size for capture-recapture data with unequal catchability. *Biometrics* 43:783–791.
- Chappell, J. 1980. Coral morphology, diversity and reef growth. *Nature* 286:249–252.
- Cohen, K., S. Finney, P. Gibbard, and J.-X. Fan. 2013. The ICS International Chronostratigraphic Chart. *Episodes* 36:199–204.
- Colwell, R. K. 2013. EstimateS: Statistical estimation of species richness and shared species from samples. Version 9. User's guide and application. <http://purl.oclc.org/estimates>.
- Colwell, R. K. R., and J. A. J. Coddington. 1994. Estimating terrestrial biodiversity through extrapolation. *Philosophical Transactions of the Royal Society B* 345:101–118.
- Darling, E. S., L. Alvarez-Filip, T. A. Oliver, T. R. McClanahan, I. M. Côté, and D. Bellwood. 2012. Evaluating life-history strategies of reef corals from species traits. *Ecology Letters* 15:1378–1386.
- de Boer, B., R. S. W. van de Wal, L. J. Lourens, and R. Bintanja. 2012. Transient nature of the Earth's climate and the implications for the interpretation of benthic $\delta^{18}\text{O}$ records. *Palaeogeography, Palaeoclimatology, Palaeoecology* 335–336:4–11.
- Devictor, V., D. Mouillot, C. Meynard, F. Jiguet, W. Thuiller, and N. Mouquet. 2010. Spatial mismatch and congruence between taxonomic, phylogenetic and functional diversity: the need for integrative conservation strategies in a changing world. *Ecology Letters* 13:1030–1040.
- Díaz, S., A. Purvis, J. H. Cornelissen, G. M. Mace, M. J. Donoghue, R. M. Ewers, P. Jordano, and W. D. Pearse. 2013. Functional traits, the phylogeny of function, and ecosystem service vulnerability. *Ecology and Evolution* 3:2958–2975.
- Edinger, E. N. 1995. Preferential survivorship of brooding corals in a regional extinction. *Paleobiology* 21:200–219.
- Edinger, E. N., and M. J. Risk. 1994. Oligocene-Miocene extinction and geographic restriction of Caribbean corals: roles of turbidity, temperature, and nutrients. *Palaios* 9:576–598.
- Ellison, A. M., E. J. Farnsworth, and R. E. Merkt. 1999. Origins of mangrove ecosystems and the mangrove biodiversity anomaly. *Global Ecology and Biogeography* 8:95–115.
- Faith, D. P., and R. Norris. 1989. Correlation of environmental variables with patterns of distribution and abundance of common and rare freshwater macroinvertebrates. *Biological Conservation* 50:77–98.
- Foggo, A., M. J. Attrill, M. T. Frost, and A. A. Rowden. 2003. Estimating marine species richness: an evaluation of six extrapolative techniques. *Marine Ecology Progress Series* 248:15–26.
- Fonseca, C. R., and G. Ganade. 2001. Species functional redundancy, random extinctions and the stability of ecosystems. *Journal of Ecology* 89:118–125.
- Foote, M. 2000. Origination and extinction components of taxonomic diversity: general problems. *Paleobiology* 26:74–102.
- Giraudel, J., and S. Lek. 2001. A comparison of self-organizing map algorithm and some conventional statistical methods for ecological community ordination. *Ecological Modelling* 146:329–339.
- Gower, J. C., and P. Legendre. 1986. Metric and Euclidean properties of dissimilarity coefficients. *Journal of Classification* 3:5–48.
- Guégan, J.-F., S. Lek, and T. Oberdorff. 1998. Energy availability and habitat heterogeneity predict global riverine fish diversity. *Nature* 391:382–384.
- Guillemot, N., M. Kulbicki, P. Chabanet, and L. Vigliola. 2011. Functional redundancy patterns reveal non-random assembly rules in a species-rich marine assemblage. *PLoS ONE* 6:e26735.
- Hall, R. 1996. Reconstructing Cenozoic SE Asia. *Geological Society of London, Special Publication* 106:153–184.
- . 2001. Cenozoic reconstructions of SE Asia and the SW Pacific: changing patterns of land and sea. Pp. 35–56 in *Faunal and floral migrations and evolution in SE Asia— Australasia*. Swets & Zeitlinger, Lisse.
- . 2002. Cenozoic geological and plate tectonic evolution of SE Asia and the SW Pacific: computer-based reconstructions, model and animations. *Journal of Asian Earth Sciences* 20:353–431.
- . 2009. Hydrocarbon basins in SE Asia: understanding why they are there. *Petroleum Geoscience* 15:131–146.
- Hall, R., M. A. Cottam, and M. E. Wilson. 2011. The SE Asian gateway: history and tectonics of the Australia–Asia collision. *Geological Society of London, Special Publication* 355:1–6.
- Hammer, Ø. 2003. Biodiversity curves for the Ordovician of Baltoscandia. *Lethaia* 36:305–313.
- Hammer, Ø., and D. A. T. Harper. 2005. *Paleontological data analysis*. Wiley-Blackwell, Malden, Mass.
- Huang, D., F. Benzoni, H. Fukami, N. Knowlton, N. D. Smith, and A. F. Budd. 2014. Taxonomic classification of the reef coral families Merulinidae, Montastraeidae, and Diploastraeidae (Cnidaria: Anthozoa: Scleractinia). *Zoological Journal of the Linnean Society* 171:277–355.
- Hutchison, C. S. 2004. Marginal basin evolution: the southern South China Sea. *Marine and Petroleum Geology* 21:1129–1148.
- Jackson, J. B., and T. P. Hughes. 1985. Adaptive strategies of coral-reef invertebrates: coral-reef environments that are regularly disturbed by storms and by predation often favor the very organisms most susceptible to damage by these processes. *American Scientist* 73:265–274.
- Jackson, J. B., A. F. Budd, and A. G. Coates. 1996. *Evolution and environment in tropical America*. University of Chicago Press, Chicago.
- Jaramillo, C. A. 2002. Response of tropical vegetation to paleogene warming. *Paleobiology* 28:222–243.
- Johnson, K. G., J. B. Jackson, and A. F. Budd. 2008. Caribbean reef development was independent of coral diversity over 28 million years. *Science* 319:1521–1523.
- Johnson, K. G., W. Renema, B. R. Rosen, and N. Santodomingo. 2015. Old data for old questions: what can the historical collections really tell us about the Neogene origins of reef-coral diversity in the Coral Triangle? *Palaios* 30:94–108.
- Johnson, K. G., M. R. Sánchez-Villagra, and O. A. Aguilera. 2009. The Oligocene–Miocene transition on coral reefs in the Falcón Basin (NW Venezuela). *Palaios* 24:59–69.
- Jurgan, H., and R. M. A. Domingo. 1989. Younger Tertiary limestone formations in the Visayan Basin, Philippines. In H. Porth and C. H. von Daniels, eds. *On the geology and hydrocarbon prospects of the Visayan Basin, Philippines*. Geologisches Jahrbuch 70:207–276. Schweizerbart, Stuttgart.
- Kay, E. A. 1996. Origin and evolutionary radiation of the Mollusca. Pp. 211–220 in J. D. Taylor, ed. *Origin and evolutionary radiation of the Mollusca*. Oxford University Press, New York.

- Keith, S., A. Baird, T. Hughes, J. Madin, and S. Connolly. 2013. Faunal breaks and species composition of Indo-Pacific corals: the role of plate tectonics, environment and habitat distribution. *Proceedings of the Royal Society of London B* 280:20130818.
- King, R. S., and C. J. Richardson. 2008. Macroinvertebrate responses to a gradient of long-term nutrient additions, altered hydroperiod, and fire. Pp. 277–319 *in* The everglades experiments: lessons for ecosystem restoration. Springer, New York.
- Klaus, J. S., B. P. Lutz, D. F. McNeill, A. F. Budd, K. G. Johnson, and S. E. Ishman. 2011. Rise and fall of Pliocene free-living corals in the Caribbean. *Geology* 39:375–378.
- Klug, C., B. Kroeger, W. Kiessling, G. L. Mullins, T. Servais, J. Frýda, D. Korn, and S. Turner. 2010. The Devonian nekton revolution. *Lethaia* 43:465–477.
- Kraft, N. J., O. Godoy, and J. M. Levine. 2015. Plant functional traits and the multidimensional nature of species coexistence. *Proceedings of the National Academy of Sciences USA* 112: 797–802.
- Kusworo, A., S. Reich, F. P. Wesselingh, N. Santodomingo, K. G. Johnson, J. A. Todd, and W. Renema. 2015. Diversity and paleoecology of Miocene coral-associated mollusks from East Kalimantan (Indonesia). *Palaos* 30:116–127.
- Laliberté, E., and P. Legendre. 2010. A distance-based framework for measuring functional diversity from multiple traits. *Ecology* 91:299–305.
- Leigh, E. G., A. O'Dea, and G. J. Vermeij. 2014. Historical biogeography of the Isthmus of Panama. *Biological Reviews* 89:148–172.
- Leleux, J., and W. Renema. 2007. Types and originals of fossil Porifera and Cnidaria of Indonesia in Naturalis. NNM Technical Bulletin 10. <http://www.repository.naturalis.nl/record/270361>.
- Lohman, D. J., M. de Bruyn, T. Page, K. von Rintelen, R. Hall, P. K. L. Ng, H.-T. Shih, G. R. Carvalho, and T. von Rintelen. 2011. Biogeography of the Indo-Australian Archipelago. *Annual Review of Ecology, Evolution, and Systematics* 42: 205–226.
- Luck, G. W., R. Harrington, P. A. Harrison, C. Kremen, P. M. Berry, R. Bugter, T. P. Dawson, F. de Bello, S. Díaz, C. K. Feld, J. R. Haslett, D. Hering, A. Kontogianni, S. Lavorel, M. Rounsevell, M. J. Samways, L. Sandin, J. Settele, M. T. Sykes, S. van den Hove, M. Vandewalle, and M. Zobel. 2009. Quantifying the Contribution of organisms to the provision of ecosystem services. *BioScience* 59:223–235.
- Lunt, P., and T. Allan. 2004. Larger Foraminifera in Indonesian biostratigraphy, calibrated to isotopic dating. Geological Research and Development Centre Museum, Workshop on Micropaleontology, Bandung, Indonesia.
- Lunt, P., and W. Renema. 2014. On the *Heterostegina*–*Tansinhokella*–*Spiroclypeus* lineage in SE Asia. *Berita Sedimentologi* 30:6–31.
- MacArthur, R. H., and E. O. Wilson. 1967. The theory of island biogeography. Princeton University Press, Princeton, N.J.
- Madin, J. S. 2005. Mechanical limitations of reef corals during hydrodynamic disturbances. *Coral Reefs* 24:630–635.
- Mantel, N. 1967. The detection of disease clustering and a generalized regression approach. *Cancer Research* 27:209–220.
- Marko, P. B., R. I. Eytan, and N. Knowlton. 2015. Do large molecular sequence divergences imply an early closure of the Isthmus of Panama? *Proceedings of the National Academy of Sciences USA* 112:E5766.
- Marshall, C. R. 1990. Confidence intervals on stratigraphic ranges. *Paleobiology* 16:1–10.
- Mayfield, M. M., S. P. Bonser, J. W. Morgan, I. Aubin, S. McNamara, and P. A. Veski. 2010. What does species richness tell us about functional trait diversity? Predictions and evidence for responses of species and functional trait diversity to land-use change. *Global Ecology and Biogeography* 19:423–431.
- McClanahan, T., M. Ateweberhan, N. Graham, S. Wilson, C. R. Sebastián, M. M. Guillaume, and J. Bruggemann. 2007. Western Indian Ocean coral communities: bleaching responses and susceptibility to extinction. *Marine Ecology Progress Series* 337:1–13.
- McMonagle, L. B. 2012. A diverse assemblage of corals from the Late Oligocene of eastern Sabah, Borneo: pre-Miocene origins of the Indo-West Pacific marine biodiversity hotspot. M.S. thesis. Durham University, Durham, U.K.
- McMonagle, L. B., P. Lunt, M. E. Wilson, K. G. Johnson, C. Manning, and J. Young. 2011. A re-assessment of age dating of fossiliferous limestones in eastern Sabah, Borneo: implications for understanding the origins of the Indo-Pacific marine biodiversity hotspot. *Palaeogeography, Palaeoclimatology, Palaeoecology* 305:28–42.
- Mihaljević, M., W. Renema, K. Welsh, and J. M. Pandolfi. 2014. Eocene-Miocene shallow-water carbonate platforms and increased habitat diversity in Sarawak, Malaysia. *Palaos* 29: 378–391.
- Moore, J. C. 2013. Diversity, taxonomical versus functional. Pp. 206–215 *in* S. A. Levin, ed. *Encyclopedia of Biodiversity*. Elsevier, Oxford.
- Morley, R. J. 2000. Origin and evolution of tropical rain forests. Wiley, Hoboken, N.J.
- Morley, R. J. 2011. Cretaceous and Tertiary climate change and the past distribution of megathermal rainforests. Pp. 1–31 *in* R. J. Morley, ed. *Tropical rainforest responses to climatic change*. Springer, Berlin.
- Morley, R. J., H. P. Morley, and P. Restrepo-Pace. 2003. Unravelling the tectonically controlled stratigraphy of the West Natuna Basin by means of palaeo-derived mid Tertiary climate changes. *Proceedings of the 29th Annual Convention of the Indonesian Petroleum Association* 1:1–24.
- Murtagh, F. 2000. Multivariate data analysis software and resources. <http://www.classification-society.org/csna/mda-sw>.
- Naeem, S., and J. P. Wright. 2003. Disentangling biodiversity effects on ecosystem functioning: deriving solutions to a seemingly insurmountable problem. *Ecology Letters* 6:567–579.
- Novak, V., N. Santodomingo, A. Röslér, E. Di Martino, J. C. Braga, P. D. Taylor, K. G. Johnson, and W. Renema. 2013. Environmental reconstruction of a late Burdigalian (Miocene) patch reef in deltaic deposits (East Kalimantan, Indonesia). *Palaeogeography, Palaeoclimatology, Palaeoecology* 374:110–122.
- O'Dea, A., and L. S. Collins. 2013. Environmental, ecological, and evolutionary change in seas across the Isthmus of Panama. *Bulletin of Marine Science* 89:769–777.
- O'Dea, A., H. A. Lessios, A. G. Coates, R. I. Eytan, S. A. Restrepo-Moreno, A. L. Cione, L. S. Collins, A. de Queiroz, D. W. Farris, and R. D. Norris. 2016. Formation of the Isthmus of Panama. *Science Advances* 2:e1600883.
- Oksanen, J., F. G. Blanchet, R. Kindt, P. Legendre, P. R. Minchin, R. O'Hara, G. L. Simpson, P. Solymos, M. H. H. Stevens, and H. Wagner. 2013. Package 'vegan.' Community ecology package. Version 2.4-1.
- Pavoine, S., J. Vallet, A.-B. Dufour, S. Gachet, and H. Daniel. 2009. On the challenge of treating various types of variables: application for improving the measurement of functional diversity. *Oikos* 118:391–402.
- Petchey, O. L., and K. J. Gaston. 2006. Functional diversity: back to basics and looking forward. *Ecology Letters* 9: 741–758.
- Porth, H., and C. H. von Daniels. 1989. On the geology and hydrocarbon prospects of the Visayan Basin, Philippines. *Geologisches Jahrbuch, Reihe B* 70:1–428.
- R Development Core Team. 2012. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
- Rachello-Dolmen, P., and D. Cleary. 2007. Relating coral species traits to environmental conditions in the Jakarta Bay/Pulau

- Seribu reef system, Indonesia. *Estuarine, Coastal and Shelf Science* 73:816–826.
- Rao, C. R. 1982. Diversity and dissimilarity coefficients: a unified approach. *Theoretical Population Biology* 21:24–43.
- Renema, W. 2007. Fauna development of larger benthic foraminifera in the Cenozoic of Southeast Asia. Pp. 179–215 in W. Renema, ed. *Biogeography, time, and place: distributions, barriers, and islands*. Springer, Dordrecht, Netherlands.
- Renema, W., D. R. Bellwood, J. C. Braga, K. Bromfield, R. Hall, K. G. Johnson, P. Lunt, C. P. Meyer, L. B. McMonagle, R. J. Morley, A. O'Dea, J. A. Todd, F. P. Wesselingh, M. E. J. Wilson, and J. M. Pandolfi. 2008. Hopping hotspots: global shifts in marine biodiversity. *Science* 321:654–657.
- Ricotta, C. 2005. Through the jungle of biological diversity. *Acta Biotheoretica* 53:29–38.
- Rosen, B., G. Aillud, F. Bosellini, N. Clack, E. Insalaco, F. Valdeperas, and M. Wilson. 2002. Platy coral assemblages: 200 million years of functional stability in response to the limiting effects of light and turbidity. *Proceedings of the Ninth International Coral Reefs Symposium, Bali* 1:255–264.
- Rosen, B. R. 1984. Reef coral biogeography and climate through the late Cainozoic: just islands in the sun or a critical pattern of islands. In P. J. Brenchley, ed. *Fossils and climate*. Geological Journal, Special Issue. 11:201–262.
- Rosenzweig, M. L. 1995. *Species diversity in space and time*. Cambridge University Press, Cambridge.
- Russel, G. J. 1998. Turnover dynamics across ecological and geological scales. Pp. 377–404. in M. L. McKinney, and J. Drake, eds. *Biodiversity dynamics: evolutionary turnover and volatility in higher taxa*. Columbia University Press, New York.
- Santodomingo, N., V. Novak, V. Petković, N. Marshall, E. Di Martino, E. L. G. Capelli, A. Roesler, S. Reich, J. C. Braga, W. Renema, and K. G. Johnson. 2015a. A diverse patch reef from turbid habitats in the middle Miocene (East Kalimantan, Indonesia). *Palaos* 30:128–149.
- Santodomingo, N., W. Renema, and K. G. Johnson. 2016. Understanding the murky history of the Coral Triangle: Miocene corals and reef habitats in East Kalimantan (Indonesia). *Coral Reefs* 35:765–781.
- Santodomingo, N., C. C. Wallace, and K. G. Johnson. 2015b. Fossils reveal a high diversity of the staghorn coral genera *Acropora* and *Isopora* (Scleractinia: Acroporidae) in the Neogene of Indonesia. *Zoological Journal of the Linnean Society* 175: 677–763.
- Scherer-Lorenzen, M., E. D. Schulze, A. Don, J. Schumacher, and E. Weller. 2007. Exploring the functional significance of forest diversity: a new long-term experiment with temperate tree species (BIOTREE). *Perspectives in Plant Ecology, Evolution and Systematics* 9:53–70.
- Sepkoski, J. J., Jr. 1975. Stratigraphic biases in the analysis of taxonomic survivorship. *Paleobiology* 1:343–355.
- Smith, A. B. 2001. Large-scale heterogeneity of the fossil record: implications for Phanerozoic biodiversity studies. *Philosophical Transactions of the Royal Society B* 356:351–367.
- Sommer, B., P. L. Harrison, M. Beger, and J. M. Pandolfi. 2014. Trait-mediated environmental filtering drives assembly at biogeographic transition zones. *Ecology* 95:1000–1009.
- Soong, K. 1993. Colony size as a species character in massive reef corals. *Coral Reefs* 12:77–83.
- Stanley, S. M., and L. A. Hardie 1998. Secular oscillations in the carbonate mineralogy of reef-building and sediment-producing organisms driven by tectonically forced shifts in seawater chemistry. *Palaeogeography, Palaeoclimatology, Palaeoecology* 144:3–19.
- Stehli, F. G., and J. W. Wells. 1971. Diversity and age patterns in hermatypic corals. *Systematic Biology* 20:115–126.
- Strauss, D., and P. M. Sadler 1989. Classical confidence intervals and Bayesian probability estimates for ends of local taxon ranges. *Mathematical Geology* 21:411–427.
- Stuart-Smith, R. D., A. E. Bates, J. S. Lefcheck, J. E. Duffy, S. C. Baker, R. J. Thomson, J. F. Stuart-Smith, N. A. Hill, S. J. Kininmonth, L. Airoidi, M. A. Becerro, S. J. Campbell, T. P. Dawson, S. A. Navarrete, G. A. Soler, E. M. A. Strain, T. J. Willis, and G. J. Edgar. 2013. Integrating abundance and functional traits reveals new global hotspots of fish diversity. *Nature* 501:539–542.
- Tager, D., J. M. Webster, D. C. Potts, W. Renema, J. C. Braga, and J. M. Pandolfi. 2010. Community dynamics of Pleistocene coral reefs during alternative climatic regimes. *Ecology* 91: 191–200.
- Tammekänd, M., O. Hints, and J. Nölvak. 2010. Chitinozoan dynamics and biostratigraphy in the Vao Formation (Darrwiliian) of the Uuga Cliff, Pakri Peninsula, NW Estonia. *Estonian Journal of Earth Sciences* 59(1), 25.
- Umbgrove, J. H. 1946. Evolution of reef corals in East Indies since Miocene time. *AAPG Bulletin* 30(1), 23–31.
- Van Valen, L. M. 1984. A resetting of Phanerozoic community evolution. *Nature* 307:50–52.
- Veron, J. E. N., and M. Stafford-Smith. 2002. *Coral ID*. Australian Institute of Marine Sciences, Townsville. <http://data.aims.gov.au/metadataviewer/uuid/3caf89c0-55b9-11dc-8d3c-00008a07204e>.
- Villéger, S., J. R. Miranda, D. F. Hernández, and D. Mouillot. 2010. Contrasting changes in taxonomic vs. functional diversity of tropical fish communities after habitat degradation. *Ecological Applications* 20:1512–1522.
- Vielle, C., M. L. Navas, D. Vile, E. Kazakou, C. Fortunel, I. Hummel, and E. Garnier 2007. Let the concept of trait be functional!. *Oikos* 116:882–892.
- Wallace, C. C., and B. R. Rosen. 2006. Diverse staghorn corals (*Acropora*) in high-latitude Eocene assemblages: implications for the evolution of modern diversity patterns of reef corals. *Proceedings of the Royal Society B* 273:975–982.
- Wannier, M. 2009. Carbonate platforms in wedge-top basins: an example from the Gunung Mulu National Park, Northern Sarawak (Malaysia). *Marine and Petroleum Geology* 26:177–207.
- Weigelt, A., J. Schumacher, C. Roscher, and B. Schmid 2008. Does biodiversity increase spatial stability in plant community biomass? *Ecology Letters* 11:338–347.
- Wilson, M. E. 2002. Cenozoic carbonates in Southeast Asia: implications for equatorial carbonate development. *Sedimentary Geology* 147:295–428.
- Wilson, M. E. J. 2008. Global and regional influences on equatorial shallow-marine carbonates during the Cenozoic. *Palaeogeography, Palaeoclimatology, Palaeoecology* 265:262–274.
- Wilson, M. E. J. 2015. Oligo-Miocene variability in carbonate producers and platforms of the Coral Triangle biodiversity hotspot: habitat mosaics and marine biodiversity. *Palaos* 30: 150–168.
- Wilson, M. E. J., and B. R. Rosen. 1998. Implications of paucity of corals in the Paleogene of SE Asia: plate tectonics or centre of origin. Pp. 165–195 in. *Biogeography and geological evolution of SE Asia*. Backhuys, Amsterdam, Netherlands.
- WoRMS Editorial Board 2016. *World Register of Marine Species*. <http://www.marinespecies.org> at VLIZ. Accessed 2016-07. doi: 10.14284/170.
- Wright, V. P., and P. M. Burgess. 2005. The carbonate factory continuum, facies mosaics and microfacies: an appraisal of some of the key concepts underpinning carbonate sedimentology. *Facies* 51(1–4), 17–23.
- Zachos, J., M. Pagani, L. Sloan, E. Thomas, and K. Billups. 2001. Trends, rhythms, and aberrations in global climate 65 Ma to present. *Science* 292:686–693.